

TECHNIQUES IN PLANT SCIENCES

No 2

Biochemical Models of Leaf Photosynthesis

S. von Caemmerer



CSIRO
PUBLISHING

National Library Cataloguing in Publication entry

von Caemmerer, S. (Susanne).

Biochemical models of leaf photosynthesis.

Bibliography.

ISBN 0 643 06379 X.

1. Botanical chemistry.

2. Photosynthesis – Measurement.

I. Title. (Series : Techniques in plant sciences ; no. 2).

572.460723

© CSIRO 2000

This book is available from:

CSIRO PUBLISHING

PO Box 1139 (150 Oxford Street)

Collingwood VIC 3066

Australia

Tel: (03) 9662 7666 Int: +(613) 9662 7666

Fax: (03) 9662 7555 Int: + (613) 9662 7555

Email: sales@publish.csiro.au

<http://www.publish.csiro.au>

Printed in Australia by Brown Prior Anderson

Contents

Preface	vii
Acknowledgments	x
1 The kinetics and regulation of rubisco	1
1.1 Introduction	1
1.2 Kinetics of fully activated rubisco	2
1.2.1 Definition of kinetic constants	2
1.2.2 Kinetics at saturating RuBP	3
1.2.3 Relative specificity, $S_{c/o}$	4
1.2.4 Rubisco kinetic constants in vitro and in vivo	6
1.2.5 RuBP-limited rate of carboxylation	10
1.2.6 High rubisco site concentrations in the chloroplast	11
1.2.7 The binding of phosphorylated compounds and other ligands	14
1.3 Activation of rubisco	16
1.3.1 Activation of rubisco in the absence of RuBP	17
1.3.2 Activation of rubisco in the presence of RuBP	17
1.3.3 The carbamylation ratio of rubisco	18
1.3.4 Rubisco activase	19
1.3.4.1 A model of activase action	20
1.3.4.2 A hyperbolic response of carbamylation to activase activity	21
1.3.4.3 Binding of other ligands	22
1.3.4.4 An unknown function of activase	22
1.4 Derivations	24
1.4.1 Derivation of rate equations for activated enzyme	24
1.4.2 The rate equations at high concentration of enzyme sites	25
1.4.3 Derivation of rate equations of activation	26
1.4.4 Carbamylated enzyme sites as a fraction of total enzyme sites	27
1.4.5 Ligands other than RuBP	28
2 Modelling C_3 photosynthesis	29
2.1 Introduction	29
2.2 A few simplifying assumptions	29
2.3 Stoichiometry of C_3 photosynthesis	30
2.3.1 Carbon stoichiometry of the PCR and PCO cycles	30
2.3.2 The carbon path from PGA to RuBP	31
2.3.3 ATP and NADPH consumption in the PCR and PCO cycles	32
2.3.4 Thylakoid reactions	32

2.3.4.1	Production of NADPH and ATP	32
2.3.4.2	Light dependence of electron transport rate	34
2.3.5	Respiration	35
2.4	Rate equations for CO ₂ assimilation	35
2.4.1	RuBP-saturated CO ₂ assimilation rate	35
2.4.2	RuBP-limited CO ₂ assimilation rate	36
2.4.3	Export-limited CO ₂ assimilation rate	38
2.4.4	Summary of rate equations	39
2.5	CO ₂ partial pressure in the chloroplast	39
2.6	Parameterization of the model	42
2.6.1	Values at 25°C	42
2.6.2	Temperature dependencies	44
2.6.2.1	Rubisco and R _d	44
2.6.2.2	Temperature dependence of J _{max}	45
2.7	The CO ₂ compensation point	47
2.7.1	Γ in the absence of R _d	48
2.7.2	Γ in the presence of R _d	48
2.7.3	Light and temperature dependence	49
2.7.4	Measurements of Γ*	50
2.8	CO ₂ response curves	51
2.8.1	CO ₂ response curves in transgenic tobacco with impaired photosynthesis	52
2.8.2	CO ₂ assimilation rate at different O ₂ partial pressures	54
2.8.3	CO ₂ assimilation rate at different irradiances	55
2.8.4	CO ₂ assimilation rate at different temperatures	56
2.8.5	The initial slope of the CO ₂ response curve, dA/dC _i	57
2.9	Light response curves	58
2.9.1	Dependence on O ₂ and CO ₂ partial pressures	58
2.9.2	Quantum yield	60
2.10	Temperature responses	62
2.11	Does the activation state of rubisco need to be incorporated into models of CO ₂ assimilation?	62
2.12	Long-term effect of environment on photosynthesis	65
2.12.1	V _{emax} and J _{max} and the transition from rubisco to electron transport limitation	65
2.12.2	Effect of growth temperature	68
2.12.3	Growth at elevated CO ₂	68
2.13	Engineering a better rubisco	70
3	Chlorophyll fluorescence and oxygen exchange during C ₃ photosynthesis	72
3.1	Introduction	72
3.2	Chlorophyll fluorescence and chloroplast electron transport	72
3.2.1	Calculating electron transport rate from chlorophyll fluorescence	73
3.2.2	Estimating electron transport rate, J _A , from CO ₂ assimilation rate	74
3.2.3	Relationship between J _f /(absI) and F _s /F _m	76
3.2.4	Calculation of mesophyll conductance to CO ₂ diffusion	78
3.2.5	Estimation of rubisco oxygenation rate from J _f and J _A	79
3.3	Oxygen exchange during C ₃ photosynthesis	80
3.3.1	Introduction	80
3.3.2	Basic equations	81
3.3.2.1	Total oxygen evolution	81

3.3.2.2	Total oxygen uptake	82
3.3.2.3	The Mehler ascorbate peroxidase (MAP) reaction	83
3.3.2.4	Net O ₂ and CO ₂ exchange	85
3.3.2.5	Estimation of rubisco carboxylation and oxygenation rates	86
3.3.3	The CO ₂ dependence of O ₂ exchange	87
3.3.4	The O ₂ dependence of O ₂ exchange	88
3.3.5	The O ₂ exchange at the compensation point	89
3.3.6	The temperature dependence of O ₂ exchange	90
4	Modelling C ₄ photosynthesis	91
4.1	Introduction	91
4.2	Basic model equations	91
4.2.1	Equations for enzyme-limited photosynthesis	94
4.2.1.1	CO ₂ assimilation rate in the bundle sheath	94
4.2.1.2	Bundle-sheath CO ₂ partial pressure	94
4.2.1.3	Bundle-sheath O ₂ partial pressure	94
4.2.1.4	The rate of PEP carboxylation	95
4.2.1.5	Quadratic expression for the enzyme-limited CO ₂ assimilation rate	95
4.2.2	Light- and electron-transport-limited photosynthesis	96
4.2.2.1	Rates of ATP and NADPH consumption	96
4.2.2.2	Partitioning of electron transport rate between C ₃ and C ₄ cycles	97
4.2.2.3	Light dependence of electron transport rate	98
4.2.2.4	Quadratic expression for electron-transport-limited CO ₂ assimilation rate	98
4.2.3	Summary of equations	99
4.3	Analysis of the model	99
4.3.1	Parameterization of the model	99
4.3.2	The model at high irradiance	101
4.3.2.1	PEP carboxylase and rubisco activity	101
4.3.2.2	Bundle-sheath conductance	102
4.3.2.3	Variation of bundle-sheath conductance, PEP carboxylase activity and rubisco with leaf age	103
4.3.2.4	CO ₂ response curves	105
4.3.2.5	Oxygen sensitivity of C ₄ photosynthesis	109
4.3.2.6	CO ₂ compensation point	110
4.3.2.7	CO ₂ diffusion from intercellular air spaces to the mesophyll cytosol	113
4.3.3	CO ₂ fixation at limiting light	116
4.3.3.1	Optimal partition of electron transport	116
4.3.3.2	Leakiness	118
4.3.3.3	Quantum yield	119
4.3.4	Modelling different decarboxylation types	120
4.3.4.1	Bundle-sheath conductance	120
4.3.4.2	O ₂ evolution in the bundle sheath	121
4.3.4.3	Energy requirements	122
5	Models of C ₃ –C ₄ intermediate photosynthesis	123
5.1	Introduction	123
5.2	Basic model equations	124
5.3	Equations for photosynthesis at high irradiance	125

5.3.1	Rubisco-limited CO ₂ assimilation rate	125
5.3.2	Net O ₂ evolution in the bundle sheath	126
5.3.3	Quadratic expression for CO ₂ assimilation rate	126
5.4	Parameterization and analysis of the model at high irradiance	127
5.4.1	Kinetic constants	127
5.4.2	Respiration	127
5.4.3	Partitioning of photorespiration	128
5.4.4	Bundle-sheath conductance	128
5.5	The glycine shuttle	129
5.5.1	Bundle-sheath rubisco activity and conductance	130
5.5.2	CO ₂ compensation point	131
5.5.3	CO ₂ and O ₂ responses of CO ₂ assimilation rate	132
5.6	The contribution of C ₄ photosynthesis	133
5.6.1	CO ₂ compensation point	134
5.6.2	CO ₂ and O ₂ responses of CO ₂ assimilation	135
5.7	Energy requirements of C ₃ –C ₄ photosynthesis	135
5.7.1	Rates of ATP and NADP consumption	135
5.7.2	Electron transport rate	137
5.7.3	Light-dependent CO ₂ assimilation rate	138
5.8	Conclusion	139
6	Concluding remarks	140
	Appendix List of symbols	141
	References	145

Preface

Increasing concerns about global climate change have revived research interests in all aspects of carbon exchange. Natural ecosystems form an important part of the global carbon balance as sinks for atmospheric CO₂. Interest in predicting net primary productivity has restored interest in leaf photosynthetic models to predict and assess changes in photosynthetic CO₂ assimilation in different environments. Photosynthetic processes of leaves have a remarkable influence on our global atmosphere. Seasonal and latitudinal variations in the carbon isotope ratio of atmospheric CO₂ relate to rubisco's preference for ¹²CO₂ rather than ¹³CO₂. The oxygen isotope composition of atmospheric CO₂ is influenced by the amount of carbonic anhydrase in the chloroplast of C₃ species and the mesophyll cytosol of C₄ species (Francey and Tans 1987; Yakir et al. 1992; Farquhar et al. 1993). This book deals exclusively with the photosynthetic processes of leaves. The models discussed are based on the underlying biochemical processes of photosynthesis and were designed to help in the interpretation of leaf gas-exchange measurements. However, because of their simplicity they have also proved valuable as submodels in a variety of other larger scale applications such as canopy photosynthesis and climate models.

At present, the techniques of genetic and molecular biology, which allow the modulation of individual plant characters, enable new questions to be asked in ecophysiology about photosynthesis and plant growth. The steady-state leaf-photosynthetic models have become an invaluable guide for the analysis of such genetic manipulation, where they are frequently used in conjunction with gas-exchange measurements to provide *in vivo* estimates of biochemical parameters.

Leaf gas-exchange measurements were first developed in the late 1950s. Penman and Schofield (1951) put the theories of diffusion of CO₂ and water vapour through stomata on a firm physical basis. Gaastra took up their ideas in the 1950s and modern analytical gas exchange is often attributed to his seminal work (Gaastra 1959). His work was a landmark because it examined CO₂ assimilation and water vapour exchange rates of individual leaves under different environmental conditions, and he distinguished between stomatal and internal resistances. Gaastra at the time concluded that the rate of CO₂ uptake was completely limited by diffusion processes at low CO₂ partial pressures and that biochemical processes became important only at high CO₂ partial pressures. Thus, gas-exchange studies focused initially on physical limitations to diffusion. Based on Gaastra's ideas, early models of leaf gas exchange had been developed as analogues of electrical resistances, and this proved useful in making a distinction between stomatal and mesophyll limitations on CO₂ assimilation. Mesophyll, or 'residual', resistance was a collective term that embodied non-stomatal diffusive factors, and included both physical and biochemical constraints.

In Australia, particularly, there was a great interest in determining the relative importance of stomatal and mesophyll resistance in limiting CO₂ assimilation rates under adverse conditions of high temperature and frequent water stresses (Bierhuizen and Slatyer 1964; Troughton and Slatyer 1969). It was not long, however, before persuasive arguments were being brought forward to show that leaf biochemistry had an important influence on the rate of CO₂ fixation, even at low CO₂ partial pressures. For example, Björkman and Holmgren (1963) made careful gas-exchange measurements of sun and shade ecotypes of *Solidago*, and noted a strong correlation between

photosynthetic rate measured at high irradiance and ambient CO_2 and the nitrogen content of leaves, and later related it to different concentrations of rubisco. Furthermore, following earlier discoveries of the O_2 sensitivity of photosynthesis, viz. an enhancement of CO_2 assimilation rate at low O_2 , Gauh and Björkman (1969) showed very elegantly that, while oxygen partial pressures did affect CO_2 assimilation rate, water vapour exchange was not affected (i.e. stomata had not responded). Clearly, the increase in CO_2 assimilation rates seen with a decrease in O_2 partial pressures could not be explained by a limitation on CO_2 diffusion. Mathematical models of leaf photosynthesis based on Gaastra's resistance equation could not accommodate this O_2 sensitivity of CO_2 assimilation. They were quickly superseded by the development of more biochemical models in the early 1970s. The discoveries by Bowes et al. (1971) that rubisco was responsible for both carboxylation and oxygenation of ribulose-1,5-bisphosphate put rubisco in the limelight. Laing et al. (1974) and Peisker (1974) were first to compare the gas exchange of leaves with the *in vitro* kinetics of rubisco.

In this book rubisco takes centre stage. Although there are many chloroplast components essential for the operation of photosynthesis, successful mathematical descriptions of photosynthesis are inevitably linked to rate equations of rubisco carboxylation and oxygenation. Chapter 1 thus deals with the kinetic properties of rubisco and these equations form the basis for the biochemical models presented in this book. In Chapter 1, *in vitro* and *in vivo* responses of rubisco are compared and analysed. Since the leaf photosynthetic models are based on rubisco's kinetic properties they have also proved a useful tool for examining the *in vivo* activity of rubisco. This is taken up in the later part of the chapter where transgenic plants with impaired photosynthetic properties are used to unravel the mysteries of *in vivo* regulation of rubisco.

Chapter 2 is a straightforward treatment of the now frequently used photosynthesis model of Farquhar et al. (1980). The chapter contains many examples of applications of the model to the analysis of transgenic plants with altered photosynthetic properties. It identifies some of the existing gaps in our knowledge, which need to be addressed because of the present need to model photosynthesis with respect to global climate change.

Chlorophyll fluorescence has emerged as a powerful, non-destructive tool for the analysis of photosynthesis and is providing insights into chloroplast electron transport rates. It is particularly useful as a field measure of photosynthetic performance and has thus stimulated considerable interest in comparisons with photosynthetic CO_2 exchange. In Chapter 3 a comparison is made between the use of measurements of chlorophyll fluorescence to estimate chloroplast electron transport rate and estimates made from gas-exchange measurements. Furthermore, the model of Farquhar et al. (1980) is used to derive rate equations for the O_2 exchange that occurs during C_3 photosynthesis.

Though the C_3 pathway of photosynthesis dominates most of the terrestrial ecosystem, the C_4 pathway of photosynthesis is important in certain agricultural and natural ecosystems and accounts for as much as 20% of global carbon fixation. The C_4 pathway is common amongst species native to tropical and subtropical grasslands. It took some very energetic grinding of C_4 leaves before rubisco was recognized as a key player in the C_4 photosynthetic pathway (Hatch 1997; Osmond 1997). It is now well recognized that the C_4 photosynthetic pathway functions as a CO_2 concentrating mechanism that provides rubisco, located in the bundle sheath, with a high CO_2 atmosphere where it can function at near CO_2 saturation with minimal oxygenase activity. This requires the cooperation between mesophyll and bundle-sheath cells, and the involvement of two cell types has complicated biochemical analysis. Here, the photosynthetic models provide an important quantitative tool to predict bundle-sheath function.

The fifth chapter discusses biochemical models of leaf photosynthesis of C_3 - C_4 intermediate species. Different biochemical variants give rise to the syndrome of C_3 - C_4 intermediacy, but all such plants have a C_4 -like leaf anatomy. C_3 - C_4 species are sometimes considered to be evolutionary intermediates between C_3 and C_4 species. The pathways revolve around efficient

refixation of photorespiratory CO_2 . Their leaf gas exchange shows a reduced oxygen sensitivity in comparison with that of C_3 species and improved photosynthetic rates at low CO_2 partial pressure. Since many of the details of these pathways remain unexplored the photosynthetic models are, of necessity, experimental. Perhaps this chapter provides the best examples of how the biochemical models presented in this book can aid in the formulation of ideas. Each photosynthesis model provides a set of hypotheses brought together in a quantitative form that can be used to design and interpret experiments.

Acknowledgments

I wish to thank C. Barry Osmond for inviting me to contribute to this series of Techniques in Plant Sciences. I have greatly enjoyed this opportunity and appreciate the encouragement and support he has provided throughout my scientific career. With his never-ending enthusiasm for science he has been a source of inspiration for me. I had the great fortune to have Graham D. Farquhar as my PhD supervisor and have been irrevocably influenced by his rigorous approach to science. I am fortunate to be able to work within the stimulating environment of the Molecular Plant Physiology Group at the Research School of Biological Science. I am indebted to John Andrews for many fascinating discussions on the mechanism and regulation of rubisco. I thank Murray Badger, John Andrews and Dean Price for the opportunity to collaborate on the analysis of transgenic plants with impaired photosynthesis. Lastly I would like to thank John R. Evans for his friendship. I am thankful for his helpful, energetic and apposite criticisms.